

## Supplementary Materials

### **A physiologically based pharmacokinetic model of doxycycline for predicting tissue residues and withdrawal intervals in grass carp (*Ctenopharyngodon idella*)**

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## **Additional method**

### **1. Cardiac output determination and calculation**

The concentration of Evans blue in each sample was detected by spectrophotometer, and then was used to calculate the area under concentration time curve (AUC) using the trapezoidal method. The semilog method was employed to better visualize the recirculation. Cardiac output was calculated for each fish using the equation described as follows:

$$Q = D / (AUC * BW) \quad (1)$$

where D is the amount of Evan blue (mg) injected into the fish, AUC (mg\*min/mL) is the area-under concentration-time curve from the time of first dye appearance to the time of dye recirculation, and BW is the body weight (kg).

### **2. Equations for absorption from the gut**

Grass carp do not have stomach. The fish gut is different from mammals, and it includes foregut, midgut, and hindgut. Food and drug absorption mainly occurs at foregut and midgut, and it occurs to some degrees in hindgut ([Smith, 1980](#)). In the present model, the gut structure was separated into two sections: one including foregut and midgut, and the other one representing hindgut. Additionally, enterohepatic circulation of DC was also considered in the model based on published studies ([Lin et al., 2015](#); [Zeng et al., 2017](#); [Li et al., 2019](#)). Oral absorption was described using the equations below.

Foregut and midgut:

$$RAI = RDOSE_{oral} - K_a * AI - K_{int} * AI + R_{bile} - K_{ehc} * AI \quad (2)$$

where RAI is the rate of change of DC in foregut and midgut (mg/h), RDOSEoral is the rate of DC following oral administration (mg/h), Ka is the absorption rate constant of DC in foregut and midgut (/h), AI is the amount of DC in foregut and midgut (mg), Kint is the transit rate constant from foregut/midgut to hindgut (/h), Rbile is the biliary excretion rate of DC (mg/h), and Kehc is the reabsorbed rate constant by the enterohepatic circulation (/h).

Hindgut:

$$RAI_h = K_{int} * AI - K_{ah} * AI_h - R_{feces} \quad (3)$$

where RAI<sub>h</sub> is the rate of change of DC in hindgut (mg/h), K<sub>ah</sub> is the absorption rate constant of hindgut (/h), AI<sub>h</sub> is the amount of DC in hindgut (mg), and R<sub>feces</sub> is the rate of change of DC in feces.

### 3. Equations for calculating partition coefficients

$$P_t = AUC_{tissue} / AUC_{plasma} \quad (4)$$

where P<sub>t</sub> is partition coefficient in non-eliminating organs, including gill, muscle+skin and liver, AUC<sub>tissue</sub> is the AUC in the tissue, and AUC<sub>plasma</sub> is the AUC in plasma.

$$P_{te} = AUC_{tissue} / (AUC_{plasma} * (1-E)) \quad (5)$$

where P<sub>te</sub> is partition coefficient in the eliminating organ kidney, and E is the renal extraction ratio calculated as renal clearance divided by the blood flow to kidney.

## Supplementary tables

**Table S1.** Summary of pharmacokinetic and tissue residue studies of doxycycline in grass carp (*Ctenopharyngodon idella*) used in the parameter calculation and model calibration

| Species               | Route | Temperature (°C) | Dose (mg/kg) | Repeat | Sex | n | Age (months) | BW (g) | Matrix        | Assay    | Ref.                             |
|-----------------------|-------|------------------|--------------|--------|-----|---|--------------|--------|---------------|----------|----------------------------------|
| Parameter calculation | IV    | 24               | 20           | 1      | NA  | 6 | 12           | 400.5  | P             | UPLC     | Present study                    |
| Parameter calculation | PO    | 24               | 20           | 1      | NA  | 6 | 12           | 450.7  | P, M, L, K, G | UPLC     | <a href="#">Xu et al., 2019b</a> |
| Model calibration     | PO    | 24               | 20           | 3      | NA  | 6 | 12           | 450.4  | P, M, L, K, G | LC-MS/MS | <a href="#">Xu et al., 2019a</a> |

Note : The abbreviations for administration route: PO, per os; IV, intravenous administration. The abbreviations for matrix: P, plasma; M, muscle+skin; L, liver; K, kidney; G, gill.

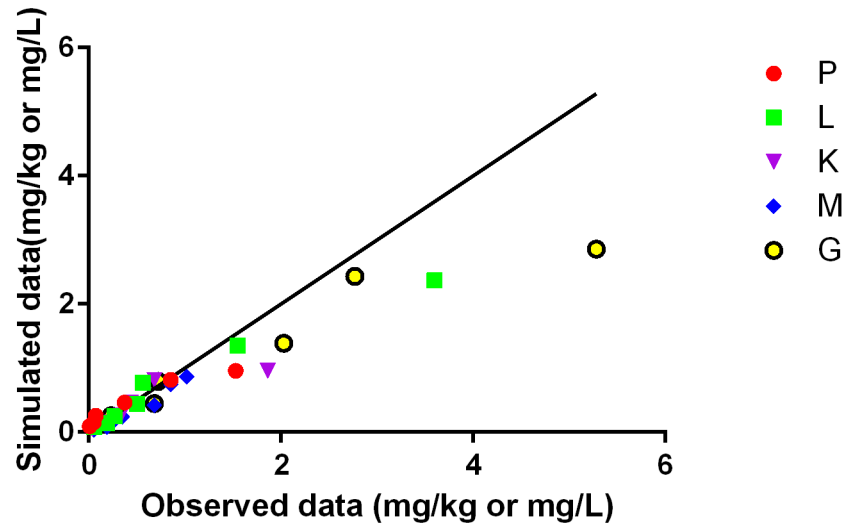
The abbreviations for determination: LC-MS/MS, liquid chromatography tandem mass spectrometry; UPLC, ultra-performance liquid chromatography. NA, not available.

**Table S2.** Pharmacokinetic parameters of doxycycline in grass carp (*Ctenopharyngodon idella*) following a single intravenous administration at 20 mg/kg

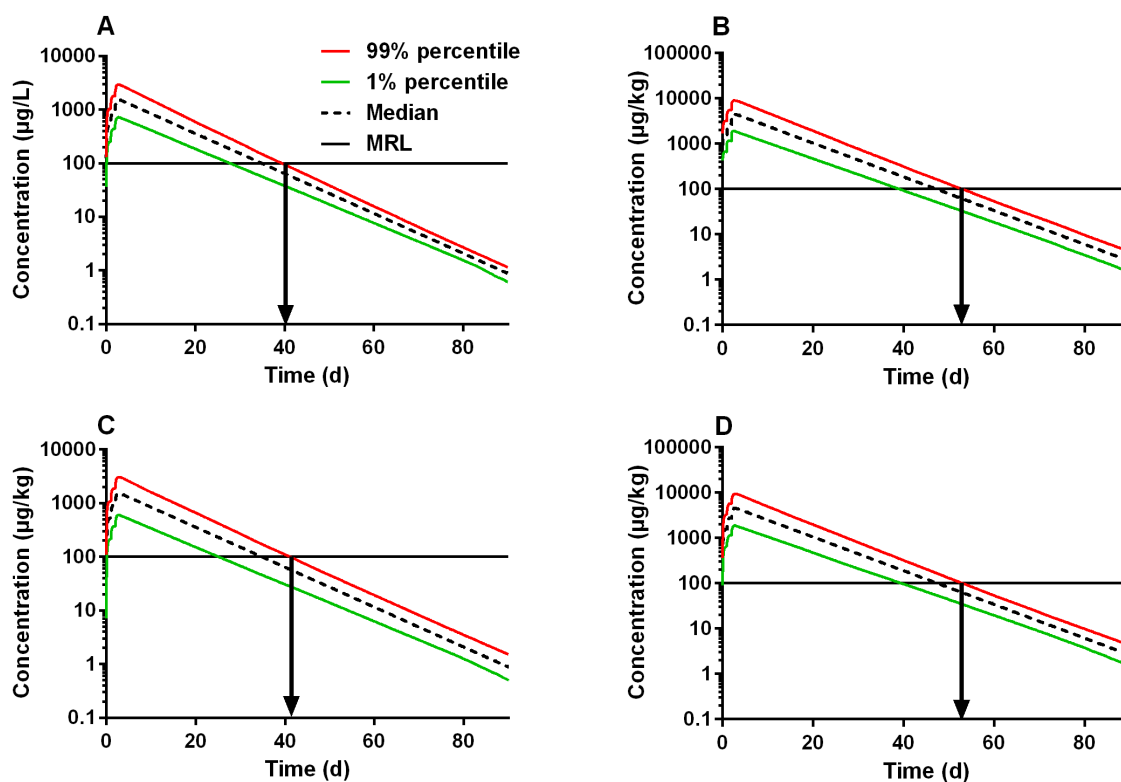
| Parameters       | Units  | Values |
|------------------|--------|--------|
| A                | mg/L   | 124.12 |
| $\alpha$         | 1/h    | 2.95   |
| B                | mg/L   | 23.32  |
| $\beta$          | 1/h    | 0.02   |
| $t_{1/2\alpha}$  | h      | 0.24   |
| $t_{1/2\beta}$   | h      | 27.75  |
| K10              | 1/h    | 0.15   |
| $t_{1/2K10}$     | h      | 4.59   |
| K12              | 1/h    | 2.33   |
| K21              | 1/h    | 0.49   |
| $AUC_{0-\infty}$ | h*mg/L | 975.78 |
| $C_{max}$        | mg/L   | 147.44 |
| $V_{ss}$         | L/kg   | 0.79   |

Note: A, zero-time blood drug concentration intercept of distribution phase;  $\alpha$ , distribution rate constant; B, zero-time blood drug concentration intercept of elimination phase;  $\beta$ , elimination rate constant;  $t_{1/2\alpha}$ , distribution half-life;  $t_{1/2\beta}$ , elimination half-life; K10, drug elimination rate constant from central compartment;  $t_{1/2K10}$ , half-life of drug leaving the body from the central compartment; K12, first-order transport rate constant from central compartment to peripheral compartment; K21, first-order transport rate constant from peripheral compartment to central compartment;  $AUC_{0-\infty}$ , area under concentration–time curve from 0 to  $\infty$ ;  $C_{max}$ , peak concentration;  $V_{ss}$ , volume of distribution at steady-state.

## Supplementary figures

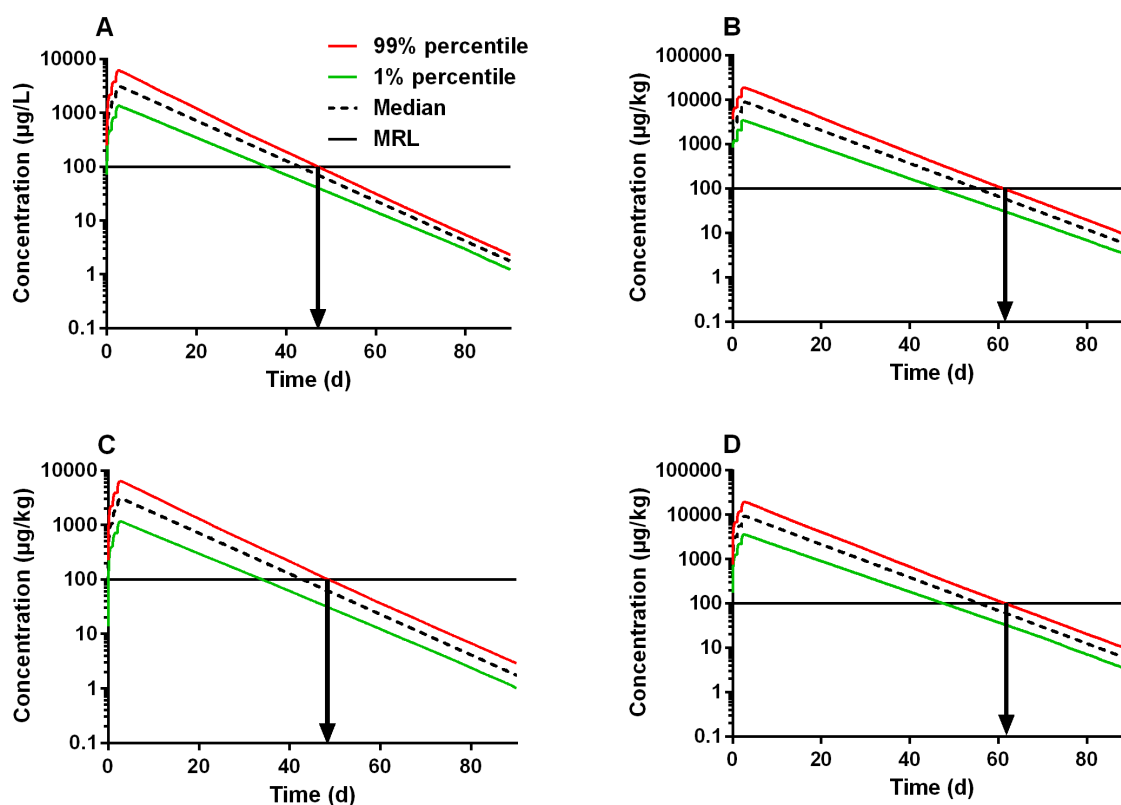


**Figure S1.** The regression analysis result between simulated data and measured data in plasma (P), liver (L), kidney (K), muscle+skin (M), and gill (G). The determination coefficient  $R^2$  value is 0.93.

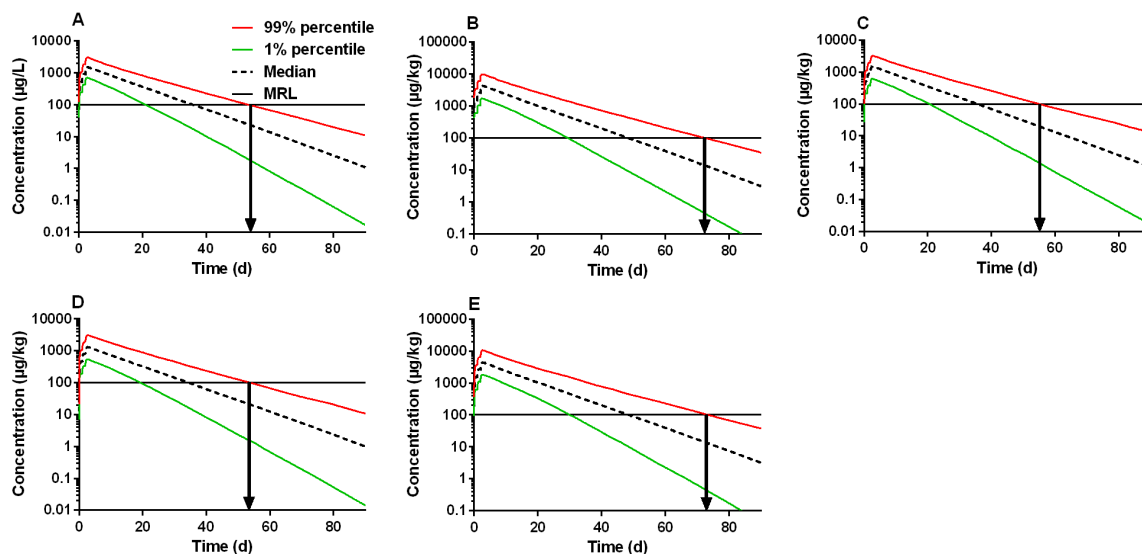


**Figure S2.** Monte Carlo simulation result for doxycycline concentrations in grass carp (*Ctenopharyngodon idella*) based on sensitive parameters by examining influence on 24-h AUC of plasma and tissues using the label dose of 20 mg/kg. The median value (black dash lines), 99<sup>th</sup> percentile (red solid lines) and 1<sup>th</sup> percentile (green solid lines) of model predictions for doxycycline concentrations in plasma (A), liver (B), kidney (C), and gill (D) of grass carp (*Ctenopharyngodon idella*) following daily oral administration at 20 mg/kg for 3 days are shown in the figure. The horizontal black line represents the maximum residue limit of 100 µg/kg for doxycycline in fish in Europe and China.

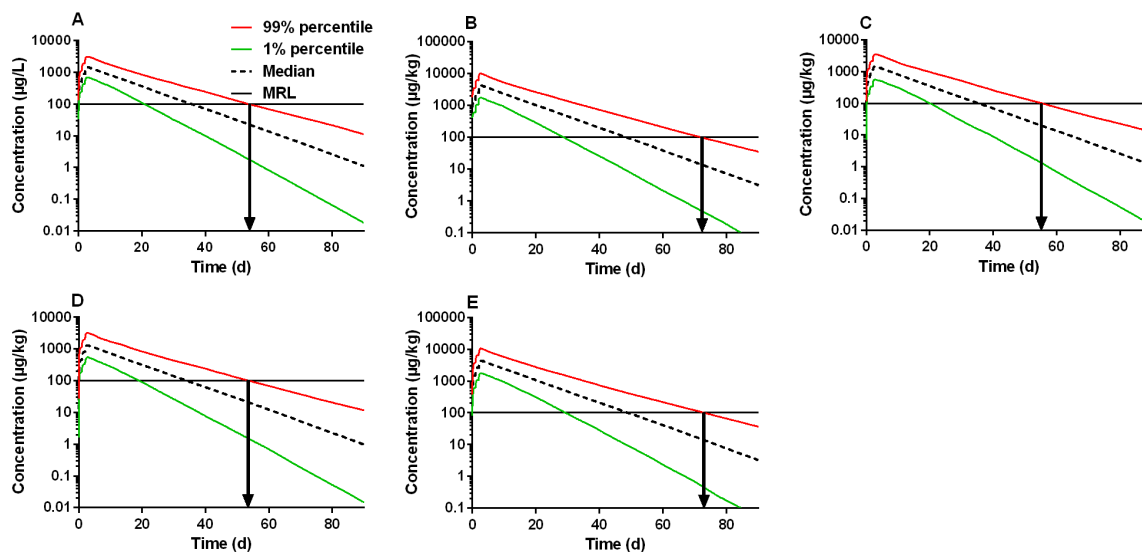




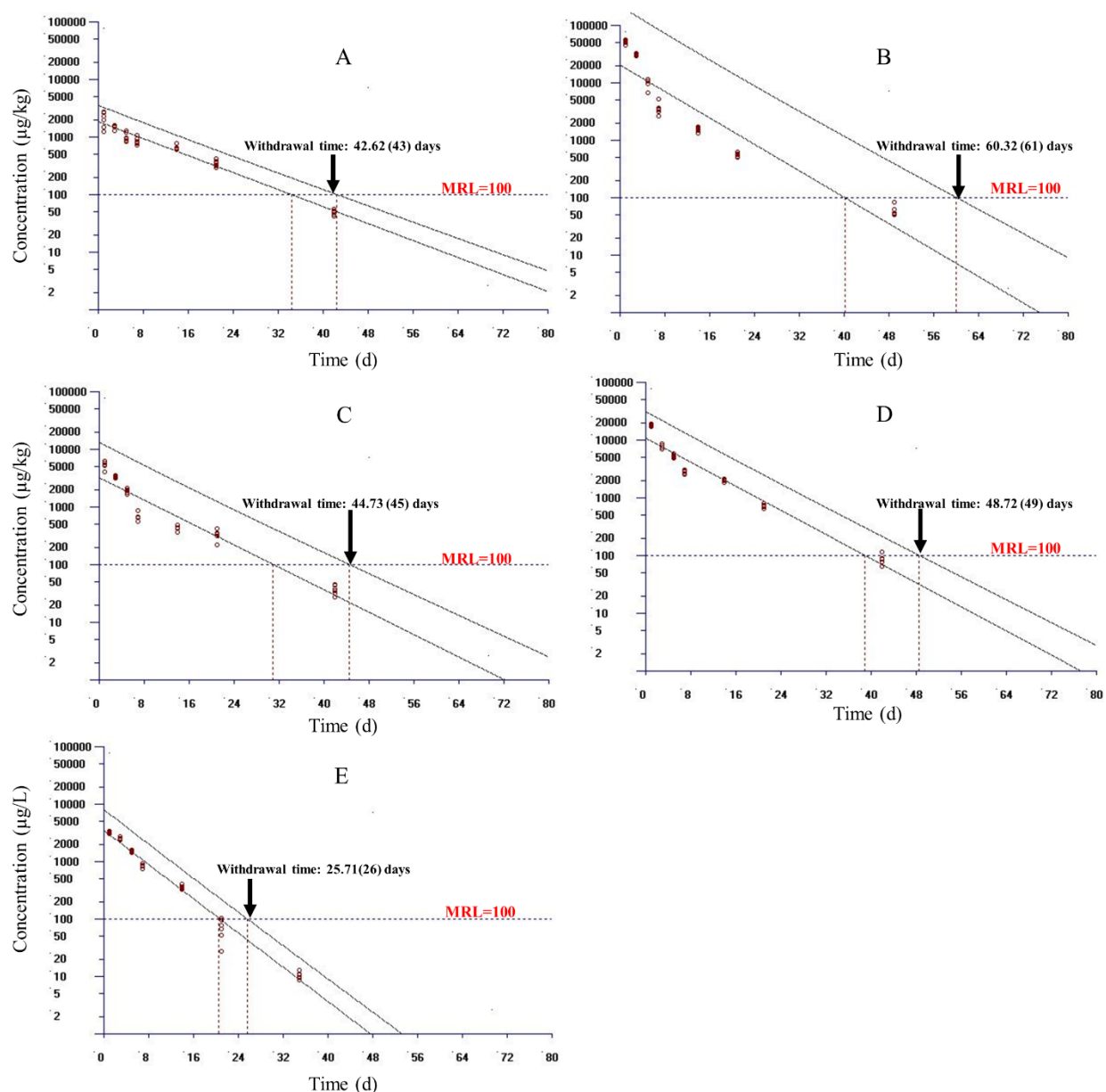
**Figure S3.** Monte Carlo simulation result for doxycycline concentrations in grass carp (*Ctenopharyngodon idella*) based on sensitive parameters by examining influence on 24-h AUC of plasma and tissues using the extra-label dose of 40 mg/kg. The median value (black dash lines), 99<sup>th</sup> percentile (red solid lines) and 1<sup>th</sup> percentile (green solid lines) of model predictions for doxycycline concentrations in plasma (A), liver (B), kidney (C), and gill (D) of grass carp (*Ctenopharyngodon idella*) following daily oral administration at 40 mg/kg for 3 days are shown in the figure. The horizontal black line represents the maximum residue limit of 100 µg/kg for doxycycline in fish in Europe and China.



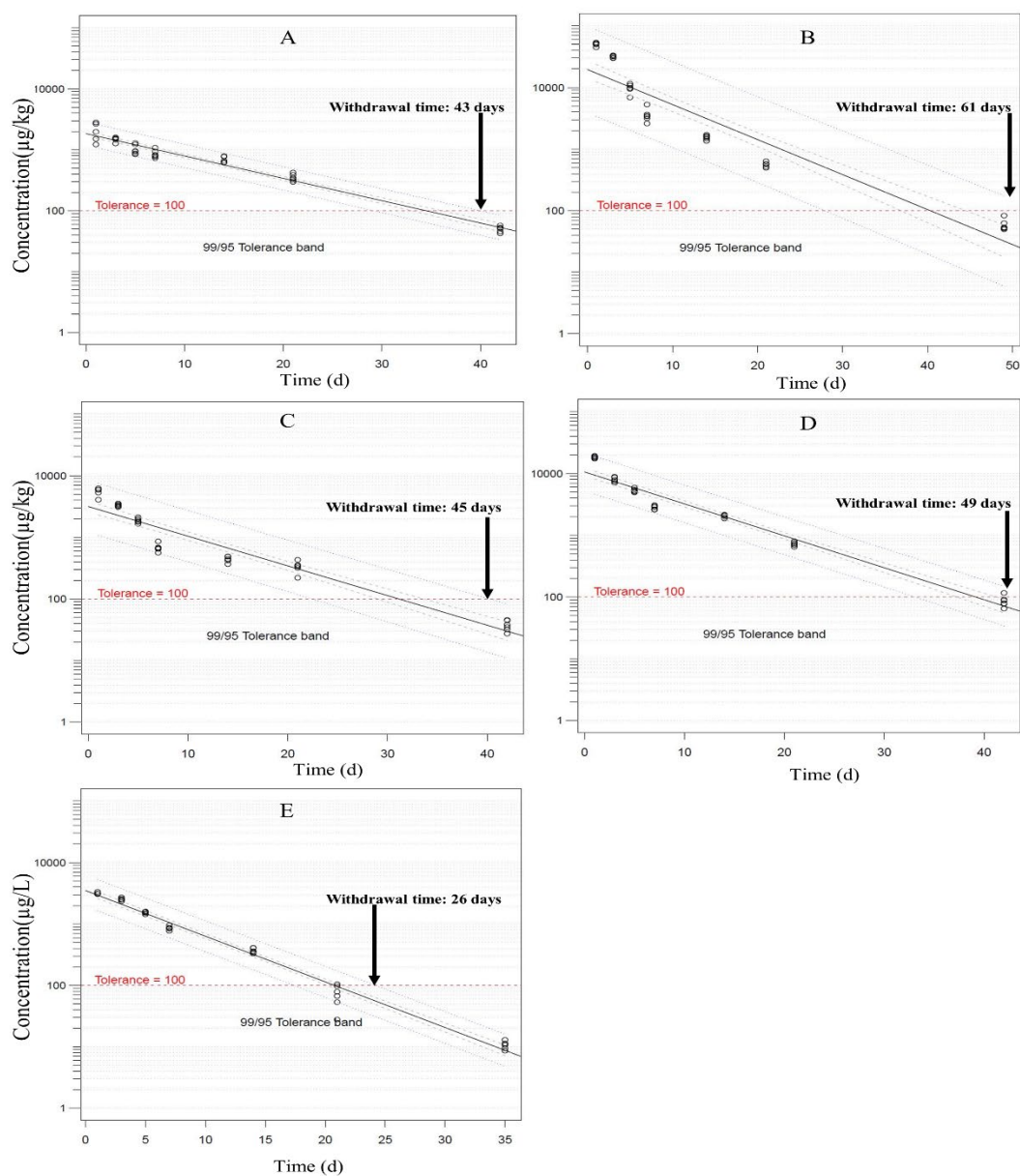
**Figure S4.** Monte Carlo simulation result for doxycycline concentrations in grass carp (*Ctenopharyngodon idella*) based on sensitive parameters by examining influence on 1008-h AUC of plasma and tissues using the label dose of 20 mg/kg. The median value (black dash lines), 99<sup>th</sup> percentile (red solid lines) and 1<sup>th</sup> percentile (green solid lines) of model predictions for doxycycline concentrations in plasma (A), liver (B), kidney (C), muscle + skin (D) and gill (E) of grass carp (*Ctenopharyngodon idella*) following daily oral administration at 20 mg/kg for 3 days are shown in the figure. The horizontal black line represents the maximum residue limit of 100 µg/kg for doxycycline in fish in Europe and China.



**Figure S5.** Monte Carlo simulation result for doxycycline concentrations in grass carp (*Ctenopharyngodon idella*) based on combined sensitive parameters by examining influence on 24-h AUC and 1008-h AUC of plasma and tissues using the label dose of 20 mg/kg. The median value (black dash lines), 99<sup>th</sup> percentile (red solid lines) and 1<sup>th</sup> percentile (green solid lines) of model predictions for doxycycline concentrations in plasma (A), liver (B), kidney (C), muscle + skin (D) and gill (E) of grass carp (*Ctenopharyngodon idella*) following daily oral administration at 20 mg/kg for 3 days are shown in the figure. The horizontal black line represents the maximum residue limit of 100 µg/kg for doxycycline in fish in Europe and China.



**Figure S6.** Estimated withdrawal times for doxycycline in grass carp (*Ctenopharyngodon idella*) using experimental data from oral administrations at 20 mg/kg for 3 days (Xu et al., 2019a) based on the EMA method using the WT 1.4 software (A for muscle+skin, B for liver, C for kidney, D for gill, and E for plasma) with a tolerance limit of 99<sup>th</sup> percentile with a 95% confidence level. MRL: maximum residue limits for doxycycline from the European Medicines Agency (EMA) (EMA, 2018). If the calculated withdrawal time was a fraction of a day, the estimated withdrawal time was rounded up to the next whole day shown in the parenthesis.



**Figure S7.** Estimated withdrawal times for doxycycline in grass carp (*Ctenopharyngodon idella*) using experimental data from oral administrations at 20 mg/kg for 3 days (Xu et al., 2019a) using FDA’s tolerance limit method coded in the “rescheme” package (A for muscle+skin, B for liver, C for kidney, D for gill, and E for plasma). The withdrawal time was calculated based on the maximum residue limit (MRL) of 100 µg/kg for DC in fish plasma and tissues with a tolerance limit of 99<sup>th</sup> percentile with a 95% confidence level.

## PBPK model code for an average individual

The model code as described below is for the physiologically based pharmacokinetic (PBPK) of doxycycline (DC) in grass carp (*Ctenopharyngodon idella*). All physiological and chemical-specific parameter values are shown in Table 1. The cardiac output and organ weight fractions were experimentally measured in the present study. The fractional blood flows in various tissues were cited from the corresponding values in rainbow trout (Law et al., 1991). The blood hematocrit in grass carp was used in the model (Yavuzcan-Yıldız and Kırkavgaç-Uzbilek, 2001). Fractional arterial plasma and venous plasma were respectively set as 0.2 and 0.8 (Lin et al., 2016). The tissue/plasma partition coefficients were calculated using pharmacokinetic data of DC in grass carp (Xu et al., 2019b). Other unknown chemical-specific parameters were obtained by model fitting to residue data of DC in grass carp (Xu et al., 2019a).

### METHOD RK4

```
STARTTIME = 0
STOPTIME = 1400
DT = 0.0025
DTOUT = 0.01
```

```
; Physiological parameters
```

```
; Blood flow rates
```

```
QCC = 3.738           ; Cardiac output (L/h/kg)
QLC = 0.181           ; Fractional blood flow to the liver
QKC = 0.102           ; Fractional blood flow to the kidney
QMC = 0.398           ; Fractional blood flow to the muscle+skin
QGC = 1               ; Fractional blood flow to the gill
QRC = 0.010           ; Fractional blood flow to the richly perfused tissues
QSC = 0.309           ; Fractional blood flow to the slowly perfused tissues
```

```
; Body weight (kg)
```

```
k = 3.896             ; The slope of the equation
Factor = 1000          ; Conversion from g to kg
Hoursinaday = 24       ; Conversion from a day to hours
BW_0 = 0.45            ; kg, The initial body weight
BW = BW_0 + (k*(TIME/Hoursinaday)) / Factor ; The equation of body weight growth
```

```
; Tissue/Organ volumes
```

```
BW = 0.450            ; Body weight (kg)
VLC = 0.004           ; Fractional liver
VKC = 0.004           ; Fractional kidney
VMC = 0.386           ; Fractional muscle+skin
VGC = 0.037           ; Fractional gill
VRC = 0.030           ; Fractional richly perfused tissues
VSC = 0.465           ; Fractional slowly perfused tissues
VartC = 0.015         ; Fractional arterial blood
VvenC = 0.059         ; Fractional venous blood
Hematocrit = 0.254    ; The blood hematocrit in grass carp
```

```
; Mass Transfer Parameters (Chemical-specific parameters)
```

```
; Partition coefficients (PC, tissue:plasma)
```

```
PL = 2.821            ; Liver:plasma PC
```

PK = 1.064 ; Kidney:plasma PC  
 PM = 0.901 ; Muscle+skin:plasma PC  
 PG = 2.981 ; Gill:plasma PC  
 PR = 2.821 ; Richly perfused tissues:plasma  
 PS = 0.901 ; Slowly perfused tissues:plasma

; Cardiac output and blood flows to tissues (L/h)

QC = QCC \* BW \* (1 - Hematocrit) ; Cardiac output  
 QL = QLC \* QC ; Liver  
 QK = QKC \* QC ; Kidney  
 QG = QGC \* QC ; Gill  
 QM = QMC \* QC ; Muscle+skin  
 QR = QRC \* QC ; Richly perfused tissues  
 QS = (1 - QLC - QKC - QMC - QRC) \* QC ; Slowly perfused tissues

; Tissue/Organ volumes

VL = VLC \* BW ; Liver  
 VK = VKC \* BW ; Kidney  
 VG = VGC \* BW ; Muscle+skin  
 VM = VMC \* BW ; Blood  
 Vart = VartC \* BW \* (1 - Hematocrit) ; Arterial plasma  
 Vven = VvenC \* BW \* (1 - Hematocrit) ; Venous plasma  
 VR = VRC \* BW ; Richly perfused tissues  
 VS = (1 - VLC - VKC - VGC - VMC - VartC - VartC - VRC) \* BW ; Slowly perfused tissues

; Kinetic constants

; Repeated oral absorption rate constants

tlen = 0.01 ; Length of oral gavage exposure (h/day)  
 tinterval = 24 ; Varied dependent on the exposure paradigm (h)  
 Tdose = 3 ; Number of dosings for multiple oral gavage  
 REPEAT [1..Tdose] = SQUAREPULSE (0 + (i - 1) \* tinterval, tlen)  
 Exposure = ARRAYSUM (REPEAT[\*])

; Oral absorption and fecal elimination rate constants

Ka = 0.007 ; /h, Absorption rate constant from foregut and midgut  
 Kah = 0.001 ; /h, Absorption rate constant from hindgut  
 Kint = 3.100e-3 ; /h, Gut transit rate constant  
 Kfeces = 0.025 ; /h, Fecal elimination rate constant

; Rate constant for the enterohepatic circulation

KehcC = 0.016 ; /h/kg

; Biliary elimination rate Constant

KbileC = 0.480 ; L/h/kg

; Metabolic rate

Kehc = KehcC \* BW ; /h

; Biliary elimination rate

Kbile = KbileC \* BW ; L/h

; Percentage of plasma protein binding, measured in the present study

PB = 0.900 ; Percentage of DC bound to plasma proteins

```

; IV infusion rate constant
Timeiv = 0.01                                ; IV injection/infusion time (h)

; Urinary elimination rate constant
KurineC = 0.019                              ; L/h/kg

; Urinary elimination rate
Kurine = KurineC * BW                        ; L/h

; Parameters for exposure scenarios
PDOSEiv = 0                                  ; mg/kg
PDOSEoral = 20                               ; mg/kg

; Dosing
DOSEiv = PDOSEiv * BW                        ; mg
DOSEoral = PDOSEoral * BW                    ; mg

; Oral dosing model
RDOSEoral = (DOSEoral / tlen) * Exposure
RAI = RDOSEoral - Ka * AI - Kint * AI + Rbile - Kehc * AI
d/dt (AI) = RAI
init AI = 0
RAIh = Kint * AI - Kah * AIh - Rfeces
d/dt (AIh) = RAIh
init AIh = 0
Rfeces = Kfeces * AIh
d/dt (Afeces) = Rfeces
init Afeces = 0
RAO = Ka * AI + Kah * AIh
d/dt (AAO) = RAO
init AAO = 0

; DC iv injection to the venous
IVR = DOSEiv / Timeiv
Riv = IVR * (1.-step(1,Timeiv))
d/dt (Aiv) = Riv
init Aiv = 0

; DC in plasma compartment
RV = QL * CVL + QK * CVK + QM * CVM + QR * CVR + QS * CVS + Riv - QC * CV
d/dt (AV) = RV
init AV = 0
CV = AV / Vven
d/dt (AUCCV) = CV
init AUCCV = 0

RA = QC * (CVG - CAfree)
d/dt (AA) = RA
init AA = 0
CA = AA / Vart
CAfree = CA * (1 - PB)

Aplasma = AV + AA

; DC in gill compartment
RG = QC * (CV - CVG)

```



```

d/dt (AG) = RG
init AG = 0
CG = AG / VG ;
CVG = AG / (VG * PG)
d/dt (AUCCG) = CG
init AUCCG = 0

; DC in liver compartment
RL = QL * (CAfree - CVL) + RAO - Rbile + Rehc
d/dt (AL) = RL
init AL = 0
CL = AL / VL ;
CVL = AL / (VL * PL)
d/dt (AUCCL) = CL
init AUCCL = 0

Rehc = Kehc * AI
d/dt (Aehc) = Rehc
init Aehc = 0

Rbile = Kbile * CVL
d/dt (Abile) = Rbile
init Abile = 0

; DC in kidney compartment
RK = QK * (CAfree - CVK) - Rurine
d/dt (AK) = RK
init AK = 0
CK = AK / VK
CVK = AK / (VK * PK)
d/dt (AUCCK) = CK
init AUCCK = 0

; Urinary excretion of DC
Rurine = Kurine * CVK
d/dt (Aurine) = Rurine
init Aurine = 0

; DC in muscle+skin compartment
RM = QM * (CAfree - CVM)
d/dt (AM) = RM
init AM = 0
CM = AM / VM
CVM = AM / (VM * PM)
d/dt (AUCCM) = CM
init AUCCM = 0

; DC in richly perfused tissue compartment
RR = QR * (CAfree - CVR)
d/dt (AR) = RR
init AR = 0
CR = AR / VR
CVR = AR / (VR * PR)

; DC in slowly perfused tissue compartment
RS = QS * (CAfree - CVS)

```

```

d/dt (AS) = RS
init AS = 0
CS = AS / VS
CVS = AS / (VS * PS)

; Mass balance
Qbal = QC - QL - QK - QM - QR - QS
Tmass = Aplasma + AL + AK + AG + AM + AR + AS + Aurine + Abile
Bal = Aiv + AAO + Aehc - Tmass

```

## Population PBPK model code

The code used to run Monte Carlo analysis is based on combined sensitive parameters by examining influence on 24-h AUC and 1008-h AUC of plasma, liver, kidney, muscle+skin, and gill.

METHOD RK4

```

STARTTIME = 0
STOPTIME = 1400
DT = 0.0025
DTOUT = 0.01

```

;Physiological parameters

;Blood flow rates

|             |                                                    |
|-------------|----------------------------------------------------|
| QCC = 3.738 | ; Cardiac output (L/h/kg)                          |
| QLC = 0.181 | ; Fractional blood flow to the liver               |
| QKC = 0.102 | ; Fractional blood flow to the kidney              |
| QMC = 0.398 | ; Fractional blood flow to the muscle+skin         |
| QGC = 1     | ; Fractional of blood flow to the gill             |
| QRC = 0.010 | ; Fractional blood flow to richly perfused tissues |
| QSC = 0.309 | ; Fractional blood flow to slowly perfused tissues |

; Body weight (kg)

|                                             |                                      |
|---------------------------------------------|--------------------------------------|
| k = 3.896                                   | ; The slope of the equation          |
| Factor = 1000                               | ; Conversion from g to kg            |
| Hoursinaday = 24                            | ; Conversion from a day to hours     |
| BW_0 = 0.45                                 | ; kg, The intial body weight         |
| BW = BW_0 + (k*(TIME/Hoursinaday)) / Factor | ; The equation of body weight growth |

;Tissue volumes

|                    |                                      |
|--------------------|--------------------------------------|
| BW = 0.450         | ; Body weight (kg)                   |
| VLC = 0.004        | ; Fractional liver                   |
| VKC = 0.004        | ; Fractional kidney                  |
| VMC = 0.386        | ; Fractional muscle+skin             |
| VGC = 0.037        | ; Fractional gill                    |
| VRC = 0.030        | ; Fractional richly perfused tissues |
| VSC = 0.465        | ; Fractional slowly perfused tissues |
| VartC = 0.015      | ; Fractional arterial blood          |
| VvenC = 0.059      | ; Fractional venous blood            |
| Hematocrit = 0.254 | ; The blood hematocrit in grass carp |

```

; Mass transfer parameters (Chemical-specific parameters)
; Partition coefficients (PC, tissue:plasma)
PL = 2.821 ; Liver:plasma PC
PK = 1.064 ; Kidney:plasma PC
PM = 0.901 ; Muscle+skin:plasma PC
PG = 2.981 ; Gill :plasma PC
PR = 2.821 ; Richly perfused tissues:plasma
PS = 0.901 ; Slowly perfused tissues:plasma

; Kinetic constants
; Repeated oral absorption rate constants
tlen = 0.01 ; Length of oral gavage exposure (h/day)
tinterval = 24 ; Varied dependent on the exposure paradigm (h)
Tdose = 3 ; Number of doses for multiple oral gavage
REPEAT[1..Tdose] = SQUAREPULSE (0+(i - 1) * tinterval, tlen)
Exposure = ARRAYSUM (REPEAT[*])

; Oral absorption and fecal elimination rate constants
Ka = 0.007 ; /h, Absorption rate constant from foregut and midgut
Kah = 0.001 ; /h, Absorption rate constant from hindgut
Kint = 3.100e-3 ; /h, Gut transit rate constant
Kfeces = 0.025 ; /h , Fecal elimination rate constant

; Rate constant for the enterohepatic circulation
KehcC = 0.016 ; /h/kg,

; Biliary elimination rate constant
KbileC = 0.480 ; L/h/kg,

; Metabolic rate constant
Kehc = KehcCm * BW ; /h

; Biliary elimination rate constant
Kbile = KbileCm * BW ; L/h

; Percentage of plasma protein binding, measured in the present study
PB = 0.900 ; Percentage of DC bound to plasma proteins

; IV infusion rate constant
Timeiv = 0.01 ; IV injection/infusion time (h)

; Urinary elimination rate constant
KurineC = 0.019 ; L/h/kg

; Urinary elimination rate
Kurine = KurineCm * BW ; L/h

; Parameters for exposure scenarios
PDOSEiv = 0 ; mg/kg
PDOSEoral = 20 ; mg/kg

; Dosing
DOSEiv = PDOSEiv * BW ; mg
DOSEoral = PDOSEoral * BW ; mg

```

```

; Variances of Parameters
BW_sd = 5.320e-2 ; Standard deviation of Body Weight
PL_sd = 5.642e-1 ; Standard deviation of PL
PK_sd = 2.128e-1 ; Standard deviation of PK
PM_sd = 1.802e-1 ; Standard deviation of PM
PG_sd = 5.926e-1 ; Standard deviation of PG
Ka_sd = 2.100e-3 ; Standard deviation of Ka
PB_sd = 2.700e-1 ; Standard deviation of PB
KehcC_sd = 4.800e-3 ; Standard deviation of KehcC
KbileC_sd = 1.440e-1 ; Standard deviation of KbileC
Kint_sd = 9.300e-4 ; Standard deviation of Kint
KurineC_sd = 5.580e-3 ; Standard deviation of KurineC

; Generation of Parameters based on Normal Distribution
init BWm = Normal(BW, BW_sd) ; Generation of the BWm based on normal distribution

; Assignment of the Values to Parameters
next BWm = BWm ; Assignment of the first created value to BWm, without this step BWm
will change at each integration time step

; Lognormal Transformation of Parameters
PL_ln = logn(PL^2/(PL_sd^2+PL^2)^0.5) ; Lognormal transformation of PL values
PL_lnsd = (logn(1+PL_sd^2/PL^2))^0.5
PK_ln = logn(PK^2/(PK_sd^2+PK^2)^0.5) ; Lognormal transformation of PK values
PK_lnsd = (logn(1+PK_sd^2/PK^2))^0.5
PM_ln = logn(PM^2/(PM_sd^2+PM^2)^0.5) ; Lognormal transformation of PM values
PM_lnsd = (logn(1+PM_sd^2/PM^2))^0.5
PG_ln = logn(PG^2/(PG_sd^2+PG^2)^0.5) ; Lognormal transformation of PG values
PG_lnsd = (logn(1+PG_sd^2/PG^2))^0.5
PB_ln = logn(PB^2/(PB_sd^2+PB^2)^0.5) ; Lognormal transformation of PB values
PB_lnsd = (logn(1+PB_sd^2/PB^2))^0.5
Ka_ln = logn(Ka^2/(Ka_sd^2+Ka^2)^0.5) ; Lognormal transformation of Ka values
Ka_lnsd = (logn(1+Ka_sd^2/Ka^2))^0.5
KehcC_ln = logn(KehcC^2/(KehcC_sd^2+KehcC^2)^0.5) ; Lognormal transformation of KehcC values
KehcC_lnsd = (logn(1+KehcC_sd^2/KehcC^2))^0.5
KbileC_ln = logn(KbileC^2/(KbileC_sd^2+KbileC^2)^0.5) ; Lognormal transformation of KbileC values
KbileC_lnsd = (logn(1+KbileC_sd^2/KbileC^2))^0.5
Kint_ln = logn(Kint^2/(Kint_sd^2+Kint^2)^0.5) ; Lognormal transformation of Kint values
Kint_lnsd = (logn(1+Kint_sd^2/Kint^2))^0.5
KurineC_ln = logn(KurineC^2/(KurineC_sd^2+KurineC^2)^0.5) ; Lognormal transformation of KurineC values
KurineC_lnsd = (logn(1+KurineC_sd^2/KurineC^2))^0.5

; Creation of Parameters based on Lognormal Distribution
init PLm = exp(Normal(PL_ln, PL_lnsd)) next PLm = PLm ; Generation of PLm based on lognormal distribution
init PMm = exp(Normal(PM_ln, PM_lnsd)) next PMm = PMm ; Generation of PMm based on lognormal distribution
init PKm = exp(Normal(PK_ln, PK_lnsd)) next PKm = PKm ; Generation of PKm based on lognormal distribution
init PGm = exp(Normal(PG_ln, PG_lnsd)) next PGm = PGm ; Generation of PGm based on lognormal distribution
init Kam = exp(Normal(Ka_ln, Ka_lnsd)) next Kam = Kam ; Generation of Kam based on lognormal distribution
init PBm = exp(Normal(PB_ln, PB_lnsd)) next PBm = PBm ; Generation of PBm based on lognormal distribution
init KehcCm = exp(Normal(KehcC_ln, KehcC_lnsd)) next KehcCm = KehcCm ; Generation of KehcCm based on
lognormal distribution
init KbileCm = exp(Normal(KbileC_ln, KbileC_lnsd)) next KbileCm = KbileCm ; Generation of KbileCm based on
lognormal distribution
init Kintm = exp(Normal(Kint_ln, Kint_lnsd)) next Kintm = Kintm ; Generation of Kintm based on
lognormal distribution

```

init KurineCm = exp(Normal(KurineC\_In, KurineC\_Insd)) next KurineCm = KurineCm ; Generation of KurineCm based on lognormal distribution

; Limit the parameter values within the lower and upper bounds

limit BWm >= 0.346

limit BWm <= 0.554

limit Kam >= 0.004

limit Kam <= 0.012

limit KehcCm >= 0.009

limit KehcCm <= 0.027

limit KbileCm >= 0.259

limit KbileCm <= 0.817

limit PLm >= 1.876

limit PLm <= 4.078

limit PKm >= 0.708

limit PKm <= 1.538

limit PMm >= 0.599

limit PMm <= 1.303

limit PGm >= 1.983

limit PGm <= 4.309

limit PBm >= 0.458

limit PBm <= 0.990

limit Kintm >= 0.002

limit Kintm <= 0.005

limit KurineCm >= 0.010

limit KurineCm <= 0.032

; Cardiac output and blood flows to tissues (L/h)

QC = QCC \* BWm \* (1 - Hematocrit)

; Cardiac output

QL = QLC \* QC

; Liver

QK = QKC \* QC

; Kidney

QG = QGC \* QC

; Gill

QM = QMC \* QC

; Muscle+skin

QR = QRC \* QC

; Richly perfused tissues

QS = QSC \* QC

; Slowly perfused tissues

; Tissue volume

VL = VLC \* BWm

; Liver

VK = VKC \* BWm

; Kidney

VG = VGC \* BWm

; Gill

VM = VMC \* BWm

; Muscle+skin

Vart = VartC \* BWm \* (1 - Hematocrit)

; Arterial plasma

Vven = VvenC \* BWm \* (1 - Hematocrit)

; Venous plasma

VR = VRC \* BWm

; Richly perfused tissues

VS = VSC \* BWm

; Slowly perfused tissues

; Oral Dosing model

RDoseOral = (DOSEoral / tlen) \* Exposure

RAI = RDoseOral - Kam \* AI - Kintm \* AI + Rbile - Kehc \* AI

d/dt (AI) = RAI

init AI = 0

RAIh = Kintm \* AI - Kah \* AIh - Rfeces

d/dt (AIh) = RAIh

init AIh = 0

Rfeces = Kfeces \* AIh

d/dt (Afeces) = Rfeces

```

init Afeces = 0
RAO = Kam * AI + Kah * AIh
d/dt (AAO) = RAO
init AAO = 0

; DC iv injection to the venous
IVR = DOSEiv / Timeiv
Riv = IVR * (1.-step(1,Timeiv))
d/dt (Aiv) = Riv
init Aiv = 0

; DC in plasma compartment
RV = QL * CVL + QK * CVK + QM * CVM + QR * CVR + QS * CVS + Riv - QC * CV
d/dt (AV) = RV
init AV = 0
CV = AV / Vven
d/dt (AUCCV) = CV
init AUCCV = 0

RA = QC * (CVG - CAfree)
d/dt (AA) = RA
init AA = 0
CA = AA / Vart ;
CAfree = CA * (1-PBm)

Aplasma = AV + AA

; DC in gill compartment
RG = QC * (CV - CVG)
d/dt (AG) = RG ;
init AG = 0
CG = AG / VG ;
CVG = AG / (VG * PGm)
d/dt (AUCCG) = CG
init AUCCG = 0

; DC in liver compartment
RL = QL * (CAfree - CVL) + RAO - Rbile + Rehc
d/dt (AL) = RL
init AL = 0
CL = AL / VL
CVL = AL/(VL* PLm)
d/dt (AUCCL) = CL
init AUCCL = 0

Rehc = Kehc * AI
d/dt (Aehc) = Rehc
init Aehc = 0

Rbile = Kbile * CVL
d/dt (Abile) = Rbile
init Abile = 0

; DC in kidney compartment
RK = QK * (CAfree - CVK) - Rurine
d/dt (AK) = RK

```

```

init AK = 0
CK = AK / VK
CVK = AK / (VK * PKm)
d/dt (AUCCK) = CK
init AUCCK = 0

; Urinary excretion of DC
Rurine = Kurine * CVK
d/dt (Aurine) = Rurine
init Aurine = 0

; DC in muscle+skin compartment
RM = QM * (CAfree - CVM)
d/dt (AM) = RM
init AM = 0
CM = AM / VM
CVM = AM / (VM * PMm)
d/dt (AUCCM) = CM
init AUCCM = 0

; DC in richly perfused tissue compartment
RR = QR * (CAfree - CVR)
d/dt (AR) = RR
init AR = 0
CR = AR / VR
CVR = AR / (VR * PR)

; DC in slowly perfused tissue compartment
RS = QS * (CAfree - CVS)
d/dt (AS) = RS
init AS = 0
CS = AS / VS
CVS = AS / (VS * PS)

; Mass balance
Qbal = QC - QL - QK - QM - QR - QS
Tmass = Aplasma + AL + AK + AG + AM + AR + AS + Aurine + Abile
Bal = Aiv + AAO + Aehc - Tmass

```

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